



## Global Council for Innovation in Rapeseed and Canola

*“Building a World community for Innovation on Rapeseed and Canola”*

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## **Editorial**

*The COVID-19 pandemics is still ongoing and all of us must adapt to this situation. Even if agricultural production has been more disturbed by climate variations in many regions than by the pandemics, the rest of the value chain has been disrupted by rapid and strong changes in consumption patterns. No restaurants means a drop in oil requirements for catering and an increase for home cooking, with different conditioning, labelling, etc... not always easy to adapt to with such short delay. No planes flying and few cars on roads mean variations on petrol markets and a drop in biodiesel needs... when cattle still need to eat oilseed meals. The flexibility of value chain is being severely tested.*

*Research life is similarly affected: closed laboratories, disturbed or cancelled programs. 2020 was a very extra-ordinary year, and 2021 is far from "normal", and we may doubt that the new normal will look like the normal of the past. However, this crisis lies positive things: new habits of work and contacts via cheap videoconferencing between distant regions have greatly facilitated the organization of the GCIRC Board and of several working meetings, the success of the Canadian Canola Week virtual event is a good example of this. Digital tools set the terms of the challenge of a broad renewal of the way of working and nature of interactions within GCIRC in the future, including the best use of our new website. Without forgetting that the sun never sets on the rapeseed empire: when it is noon in Europe, it is 10 p.m. in Australia and 5 a.m. in Western Canada. After the first experiences in 2020, we have in 2021 to learn how to optimize the use of these technologies for a global community like the GCIRC.*

*Etienne Pilorgé*

*GCIRC General Secretary*

## **Activity/ News of the association:**

### **GCIRC 2021 Technical Meeting**

The GCIRC Technical Meeting and the GCIRC General Assembly were planned to be organized in 2021 in Poznan, Poland, historical site of Rapeseed research in Europe. The COVID-19 19 crisis makes the situation too uncertain to organize a physical on site meeting this year. The GCIRC board will examine the alternative solutions beginning of March, and GCIRC members will be informed of plans as soon as possible.

## Welcome to New GCIRC members

**Ward TOMA** from Alberta Canola joined GCIRC last November.

You may visit his personal page on the GCIRC website directory, to better know his fields of interest.

*We take this opportunity to remind all members that they can modify their personal page, especially indicating their fields of interest in order to facilitate interactions.*

## Value chains and regional news

- **USA: Cibus Announces Successful Field Trials for Disease Tolerant Trait to sclerotinia in Canola (press release on January 12, 2021)**

Cibus announced that « its first field trials have confirmed greenhouse results of a non-GMO trait that provides tolerance to white mold (*Sclerotinia*). This milestone represents a major step in Cibus' mission to breed a durable resistant plant with its Rapid Trait Development System (RTDS™), as well as a breakthrough for a complex challenge that GMO and traditional technologies have been unable to fully address. This trait is among Cibus' 14 trait products that the US Department of Agriculture, under its "Am I Regulated" process, has recently ruled were not subject to regulation under 7 CFR Part 340 and instead are regulated in the same manner as traits developed using conventional breeding practices. » Read more at <https://www.cibus.com/press-release.php?date=011221> . Reported by D. Gouache, Terres Inovia.

- **USA: missions of US Canola Association slightly evolve**

During its autumn meeting, the USCA Board approved a slightly revised mission statement and updated communications strategy focused on increasing U.S. canola production. The new mission is “to increase domestic canola production to meet growing demand for healthy oil, meal and protein by promoting policies and conditions favorable to growing, marketing, processing, and using U.S. canola.” Read more on <https://www.uscanola.com/newsletter/canola-quick-bytes-december-2020/>

- **Canada: Canola Week 2020 online**

For the first time, due to the COVID-19 pandemics, the Canola Week 2020 was an online event on December 1-3. It had 650 registrants, with up to 400 participating at one time. Registration data show that 51 per cent were from research and development, 21 per cent were agronomists, 12 per cent from industry, 10 per cent producers and 6 per cent from sales. Geographically, 92 per cent were from Canada (almost all from the Prairie provinces) and 6 per cent were from the United States. The experience is quite successful, despite its limits regarding social aspects and networking.

A full reporting of the 2020 campaign and challenges for canola production in the Prairies was presented (see: <https://www.canolacouncil.org/canola-watch/2020/12/16/highlights-from-canola-week-2020/> ).

Regarding Canola economy, the COVID-19 raised interrogations on the capacity of the supply chain to remain fluid and the diversification of outlets. The Canadian industry expects the biofuel market to be a driver of growth, partly under the influence of the US, in particular California, which is also a large buyer of oilcake, and probably Europe. Market access for sustainable biofuels is then a key challenge for the coming years, for both the domestic and export markets, leading to look for "low-carbon canola", "the Clean Fuel" standard could induce a "renaissance" of up to 1.5MT. Nevertheless, the main objectives remain the food market with restaurants and industries, and yield enhancement at farms level.

Some highlights in science and innovation presentations:

- In agronomy, the AAFC presented studies on canola greenhouse gas emissions: in the conditions of the Canadian Prairies, giving a range of variation of N<sub>2</sub>O emissions from 0,16 to 0,8kgN<sub>2</sub>O-N per 100kg of N fertilizer, below the default IPCC standard (1.0). Another AAFC communication explored the use of satellite remote sensing to assess the risk of sclerotinia using risk grid criteria: mobilization of historical data on the crops in fields, observation of areas where the soil remains wet, crop phenology by cross-referencing observations with crop models, all leading to a prototype disease risk assessment tool, the DiRT, "which does not identify and predict the existence of the disease but provides a framework of geolocalized data to test existing models, hypotheses and develop new prediction models". Interesting idea for R&D, and perhaps practical applications in the future.
- Regarding genetics and breeding, the University of Alberta (Gavin Chen) worked on the development of a GMO canola rich in punicic acid (C18:3 / delta9cis-11trans-13cis), with promising results at lab level. Punicic acid is naturally present in pomegranate seeds (65% of GA), and is believed to have anticancer activity on prostate cancer, used in food supplements and cosmetics. A very interesting presentation was done by Sally MacKenzie (Pennsylvania State University) on the introduction of epigenetics in plant breeding, based on the possibility of creating phenotypic variability and a memory of the stresses endured. The work was carried out on tomato, soybean, sorghum, millet, tobacco, and canola, and published in May 2020: <https://doi.org/10.1038/s41467-020-16036-8>
- On valorization aspects, let us highlight a presentation by Christopher Hald, Technical University of Munich, on the role of Kaempferol-glycoside as the origin of the bitterness of proteins from rapeseed using a "sensomics analysis" approach. The cruciferin fraction of rapeseed proteins is predominantly bitter, astringent, and sour, whereas napins are predominantly astringent. HPLC analyses enable the Kaempferol-glycoside to be identified. The levels in rapeseed are then measured, and the impact on bitterness confirmed by adding the compound to milk casein. The authors conclude on the technological and genetic pathways to reduce bitterness.

- **Canada: highlights on Canola protein challenges by Johann F. Tergesenn President and CEO of Burcon**

The USCA Blog reported on October 28<sup>th</sup>, 2020, J.F. Fergensen's vision of the recent movements and perspectives regarding the use of vegetable proteins, notably canola's, in food industries. New added value for rapeseed/canola products is good news for the sector, in the hope that part of the added value will go back up the value chain. See <https://www.uscanola.com/news-views/usca-blog/canola-protein-for-foods-and-beverages-coming-soon/>.

- **Gene editing can modify canola architecture**

A study lead by researchers at the University of Calgary used gene editing to modify canola's genes, producing shorter plants with more branches and flowers which could potentially increase the crop's yield, the university says in a news release on Feb. 1. Read more on <https://seedworld.com/new-study-finds-possible-higher-yielding-canola/> Source: SeedWorld.com, reported by W. Keller.

For the original publication in Plant Biotechnology Journal last November: see section Scientific news/Breeding: Stanic M et al.

- **UK: London launches consultation to review its regulations on gene editing (source AgraPresse, January 13, 2021)**

The British Agriculture Minister, George Eustice, announced on January 7<sup>th</sup> the launch of a consultation on gene editing techniques in agriculture, with a view to a less stringent regulation than the European GMO regulation. The UK minister promises more flexible regulation than the one in force in Europe since the European Court of Justice (ECJ) associated with the GMO directive the other new genome-modification technologies (NBTs) that emerged after the GMO directive was adopted in 2001.

- **China: rapeseed production stable around 13,5MMT**

"Marketing Year 20/21 rapeseed production is forecast at 13.5 MMT, higher than the USDA official and US Foreign Agricultural Service China estimates for the previous year based on a slightly higher marketing price due to decreased imports. The China's National Grain and Oils Information Center forecast for 20/21 rapeseed production is 13.9 MMT, basically unchanged from its production estimate for 19/20." Source [USDA Oilseeds and products update Oct 2020](https://usda.mkt.usda.gov/oilseeds-products/2020-10-oilseeds-products-update)

- **India: high demand for mustard oil**

In India during cropping season 2020-21, the area under rapeseed and mustard, the main winter oilseeds have increased by 6.7% to 7.3 million hectares. This is the highest ever area cultivated under rabi oilseeds. Strong demand for mustard oil during the COVID-19 pandemic and higher crushing may also contribute to higher sowing this year. Mustard oil is considered to be an immunity booster. Mustard prices rose to a record high of 6,400 rupees per 100 kg (equal to 727 euro per metric ton) during last months. Source: P. Sharma, ICAR.

- **High prices on oilseeds, oils, and meals markets**

World prices of basic foodstuffs soared in January to their highest level since July 2014, driven by rising prices of cereals, oils and sugar, the FAO announced on February 4<sup>th</sup>. The monthly rise in vegetable oils was 5.8%, its highest level since May 2012, with a "staggering increase" of 14.5% compared to October. The rise in vegetable oils would be due to lower-than-expected palm oil production in Indonesia and Malaysia due to excessive rains and a persistent shortage of migrant workers. Soybean prices have also soared over the past 8 months due to reductions in export availability, strikes in Argentina, and strong demand from China. The two price drivers of the oilseed complex are therefore under pressure.

A presentation by the European Commission dated January 28<sup>th</sup> summarizes these trends: Global oilseed production is expected to be further reduced to 594 million tons on unfavorable weather in South America and Black Sea region. Prices of Oilseeds continued to increase sharply at the end of 2020 and start of 2021 on unfavorable conditions in producing regions and strong demand. Lately the trend slowed down on news of beneficial rains in South America and hopes of steep increase in acreage and production next season, driven by current high prices. Rapeseed prices also slowed down after recent sharp increases and seem to stabilize at high levels. Ukraine remains the most competitive origin for EU market. Global rapeseed production seems stable at 69 million tons.

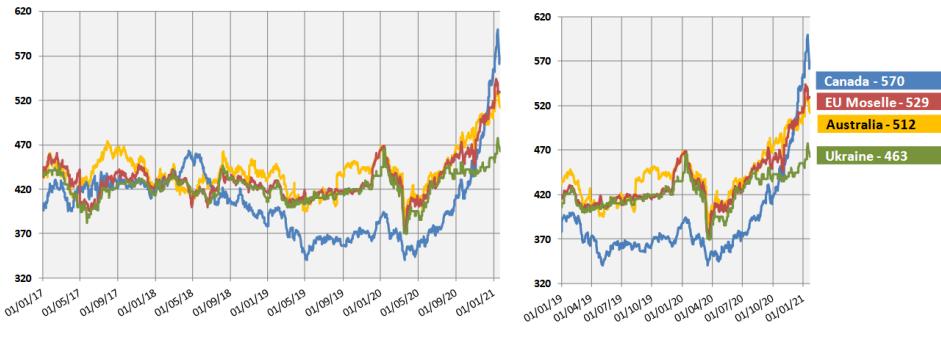
See FAO: <http://www.fao.org/worldfoodsituation/foodpricesindex/en/>

See USDA: <https://apps.fas.usda.gov/psdonline/circulars/oilseeds.pdf>

Seed "Chart of the week (02 2021) on UFOP website: Soybean prices drove oilseed meal prices:  
<https://www.ufop.de/english/news/chart-week/>

See EU Commission: <https://circabc.europa.eu/sd/a/215a681a-5f50-4a4b-a953-e8fc6336819c/oilseeds-market%20situation.pdf> ):

## World export prices for rapeseed – (USD/tonne)



Source: International Grains Council

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- **Europe: BASF and VanderSat: new tool for rapeseed monitoring:**

Germany's BASF and Holland's VanderSat have announced on January 25<sup>th</sup> that they have signed a commercial agreement to provide daily images of biomass without interference from cloud cover. The VanderSat tool, based on passive microwave technology, will be grafted onto the decision support tool for monitoring the health of rapeseed, wheat and winter barley designed by Xarvio (BASF Digital farming) to enable farmers to "accurately monitor crop growth". Tests were successfully conducted in Germany, Ukraine, the United Kingdom, Canada, and Brazil during the 2019-20 campaign. They are based on three different satellite technologies: a VanderSat method, based on passive microwave technology, active microwaves, and optical images from the European agency ESA's Sentinel satellites.

The tool, which will be operational by the end of March, will be offered first in North and Latin America, and then beyond in the course of 2021. Only the United States, Canada, Brazil, Argentina, Germany, and Ukraine are currently targeted. Source AgraPresse, January 27, 2021

## Scientific news

### Publications:

#### BREEDING

**Focus:** Reported by Wilf Keller, former GCIRC President, this article gives an overview of the use of genetic and gene technologies, for a better understanding of what is at stake with these technologies for rapeseed/canola future.

Ton, L. B., Neik, T. X., & Batley, J. (2020). The Use of Genetic and Gene Technologies in Shaping Modern Rapeseed Cultivars (*Brassica napus* L.). *Genes*, 11(10), 1161.  
<https://doi.org/10.3390/genes11101161>

Song, J. M., Liu, D. X., Xie, W. Z., Yang, Z., Guo, L., Liu, K., ... & Chen, L. L. (2020). BnPIR: *Brassica napus* Pan-genome Information Resource for 1,689 accessions. *Plant Biotechnology Journal*.  
<https://doi.org/10.1111/pbi.13491>

Chen, X., Tong, C., Zhang, X., Song, A., Hu, M., Dong, W., ... & Zhang, L. (2020). A high-quality *Brassica napus* genome reveals expansion of transposable elements, subgenome evolution and disease resistance. *Plant Biotechnology Journal*. <https://doi.org/10.1111/pbi.13493>

Arimura, S. I., Ayabe, H., Sugaya, H., Okuno, M., Tamura, Y., Tsuruta, Y., ... & Tsutsumi, N. (2020). Targeted gene disruption of ATP synthases 6-1 and 6-2 in the mitochondrial genome of *Arabidopsis thaliana* by mitoTALENs. *The Plant Journal*. <https://doi.org/10.1111/tpj.15041>

Horvath, D. P., Stamm, M., Talukder, Z. I., Fiedler, J., Horvath, A. P., Horvath, G. A., ... & Anderson, J. V. (2020). A New Diversity Panel for Winter Rapeseed (*Brassica napus*, L.) Genome-Wide Association Studies. *Agronomy*, 10(12), 2006. <https://doi.org/10.3390/agronomy10122006>

Yang, H., Bayer, P. E., Tirnaz, S., Edwards, D., & Batley, J. (2021). Genome-Wide Identification and Evolution of Receptor-Like Kinases (RLKs) and Receptor like Proteins (RLPs) in *Brassica juncea*. *Biology*, 10(1), 17. <https://doi.org/10.3390/biology10010017>

Horvath, D. P., Zhang, J., Chao, W. S., Mandal, A., Rahman, M., & Anderson, J. V. (2020). Genome-Wide Association Studies and Transcriptome Changes during Acclimation and Deacclimation in Divergent *Brassica napus* Varieties. *International journal of molecular sciences*, 21(23), 9148. <https://doi.org/10.3390/ijms21239148>

Nikzad, A., Kebede, B., Bhavikkumar, J., & Rahman, H. (2020). Study of the genetic structure of a *Brassica napus* canola population derived from six interspecific crosses of *B. napus* × *B. oleracea*. *Canadian Journal of Plant Science*, (ja). <https://doi.org/10.1139/CJPS-2020-0059>

Zhou, X. T., Jia, L. D., Duan, M. Z., Chen, X., Qiao, C. L., Ma, J. Q., ... & Li, J. N. (2020). Genome-wide identification and expression profiling of the carotenoid cleavage dioxygenase (CCD) gene family in *Brassica napus* L. *Plos one*, 15(9), e0238179. <https://doi.org/10.1371/journal.pone.0238179>

- Tong, J., Walk, T.C., Han, P. et al. Genome-wide identification and analysis of high-affinity **nitrate transporter 2 (NRT2) family genes** in rapeseed (*Brassica napus L.*) and their responses to various stresses. BMC Plant Biol 20, 464 (2020). <https://doi.org/10.1186/s12870-020-02648-1>
- Matuszczak, M., Spasibionek, S., Gacek, K. et al. Cleaved amplified polymorphic sequences (CAPS) marker for identification of two mutant alleles of the rapeseed BnaA.FAD2 gene. Mol Biol Rep 47, 7607–7621 (2020). <https://doi.org/10.1007/s11033-020-05828-2>
- Kamiński, P., Marasek-Ciolakowska, A., Podwyszyńska, M., Starzycki, M., Starzycka-Korbas, E., & Nowak, K. (2020). Development and Characteristics of **Interspecific Hybrids** between *Brassica ole-racea L.* and *B. napus L.* Agronomy, 10(9), 1339. <https://doi.org/10.3390/agronomy10091339>
- Daurova, A., Daurov, D., Volkov, D., Zhapar, K., Raimbek, D., Shmekova, M., & Zhambakin, K. (2020). Doubled haploids of **interspecific hybrids** between *Brassica napus* and *Brassica rapa* for canola production with valuable breeding traits. OCL, 27, 45. <https://doi.org/10.1051/ocl/2020041>
- Nikzad, A. (2020). Potential of the **Brassica oleracea gene pool** for the improvement of spring *B. napus canola*. (PhD thesis) <https://era.library.ualberta.ca/items/c29af9c7-ccf0-456e-8613-c8c4690d8585>
- Neik, T. X., Amas, J., Barbetti, M., Edwards, D., & Batley, J. (2020). Understanding **host-pathogen interactions** in *brassica napus* in the omics era. Plants, 9(10), 1336. <https://doi.org/10.3390/plants9101336>
- Poveda, J., Francisco, M., Cartea, M. E., & Velasco, P. (2020). Development of **Transgenic** *Brassica* Crops against **Biotic Stresses** Caused by Pathogens and Arthropod Pests. Plants, 9(12), 1664. <https://doi.org/10.3390/plants9121664>
- Cantila, A. Y., Saad, N. S. M., Amas, J. C., Edwards, D., & Batley, J. (2021). Recent Findings Unravel Genes and Genetic Factors Underlying ***Leptosphaeria maculans*** Resistance in *Brassica napus* and Its Relatives. International Journal of Molecular Sciences, 22(1), 313. <https://doi.org/10.3390/ijms22010313>
- Pröbsting, M. (2020). Application of CRISPR-Cas9 genome editing systems for improving oilseed rape (*Brassica napus*) disease resistance against ***Verticillium longisporum*** (Doctoral dissertation University of Kiel). [https://macau.uni-kiel.de/receive/macau\\_mods\\_00000442?lang=en](https://macau.uni-kiel.de/receive/macau_mods_00000442?lang=en)
- Jiang, J., Fredua-Agyeman, R., Hwang, S. F., & Strelkov, S. E. (2021). Differentially expressed genes in canola (*Brassica napus*) during infection by virulent and avirulent ***Plasmoidiophora brassicae*** pathotypes. Plant Pathology. <https://doi.org/10.1111/ppa.13267>
- Summanwar, A. (2020). Transcriptomic responses in spring canola carrying **clubroot resistance** introgressed from rutabaga or “Mendel”. PhD thesis University of Alberta. <https://era.library.ualberta.ca/items/2d2247a9-6204-469e-a585-5aed27b8f57f>
- Shaikh, R., Farid, M., & Rahman, H. (2020). Inheritance of resistance to the newly identified ***Plasmoidiophora brassicae*** pathotypes in *Brassica napus L.* Canadian Journal of Plant Pathology, 1-11. <https://doi.org/10.1080/07060661.2020.1823483>

- Zhou, Q. (2020). Comparative transcriptome analysis of rutabaga (*Brassica napus*) cultivars in response to ***Plasmidiophora brassicae***. (Master dissertation). <https://era.library.ualberta.ca/items/6ec878a1-5585-46b4-a75d-b3092e4a7589>
- Li, X., Xiang, F., Zhang, W., Yan, J., Li, X., Zhong, M., ... & Zhao, X. (2020). Characterization and Fine Mapping of a New Dwarf Mutant in *Brassica Napus*. [REFERENCE](#)
- Calderwood, A., Lloyd, A., Hepworth, J., Tudor, E. H., Jones, D. M., Woodhouse, S., ... & Morris, R. J. (2020). Total FLC transcript dynamics from divergent parologue expression explains **flowering diversity** in *Brassica napus*. *New Phytologist*. <https://doi.org/10.1111/nph.17131>
- Chai, L., Zhang, J., Li, H., Zheng, B., Jiang, J., Cui, C., & Jiang, L. (2020). Investigation for a **multi-silique trait** in *Brassica napus* by alternative splicing analysis. *PeerJ*, 8, e10135. <https://doi.org/10.7717/peerj.10135>
- Stanic, M., Hickerson, N. M., Arunraj, R., & Samuel, M. A. (2020). Gene-editing of the strigolactone receptor BnD14 confers promising **shoot architectural changes** in *Brassica napus* (canola). *Plant Biotechnology Journal*. <https://doi.org/10.1111/pbi.13513>
- Liersch, A., Bocianowski, J., Nowosad, K., Mikołajczyk, K., Spasibionek, S., Wielebski, F., ... & Bartkowiak-Broda, I. (2020). Effect of **Genotype × Environment Interaction** for Seed Traits in Winter Oilseed Rape (*Brassica napus* L.). *Agriculture*, 10(12), 607. <https://doi.org/10.3390/agriculture10120607>
- Xu, P., Wang, X., Dai, S., Cui, X., Cao, X., Liu, Z., & Shen, J. (2020). The multilocular trait of rapeseed is ideal for **high-yield breeding**. *Plant Breeding*. <https://doi.org/10.1111/pbr.12880>
- Xie, Y., Xu, J., Tian, G. et al. Unraveling **yield-related traits** with QTL analysis and dissection of QTL × environment interaction using a high-density bin map in rapeseed (*Brassica napus*. L.). *Euphytica* 216, 171 (2020). <https://doi.org/10.1007/s10681-020-02708-5>
- Cui, Y., Zeng, X., Xiong, Q., Wei, D., Liao, J., Xu, Y., ... & Qian, W. (2020). Combining QTL and co-expression analysis allowed identification of new candidates for **oil accumulation** in rapeseed. *Journal of Experimental Botany*. <https://doi.org/10.1093/jxb/eraa563>
- Islam, M. A. U., Nupur, J. A., & Robin, A. H. K. (2020). Trait Association, Genetic Analyses and **Fatty Acid Profiles** in Oilseed Producing Rapeseed-Mustard (*Brassica spp.*) Genotypes. *Plant Breeding and Biotechnology*, 8(4), 316-326. <https://doi.org/10.9787/PBB.2020.8.4.316>
- Cegielska-Taras, T., Dobrzycka, A., Bartkowiak-Broda, I., Gacek, K., Wolko, J., Bocianowski, J., & Szala, L. (2020). Genetic variation of traits affecting meal quality in **black × yellow seeded doubled haploid** population of winter oilseed rape. <https://doi.org/10.15159/ar.20.209>
- Sashidhar, N. (2020). Random and targeted CRISPR-Cas mutagenesis to create oilseed rape with **reduced seed phytic acid content** (Doctoral dissertation). University of Kiel. [https://macau.uni-kiel.de/receive/macau\\_mods\\_00000410](https://macau.uni-kiel.de/receive/macau_mods_00000410)
- Wolko, J., Dobrzycka, A., Bocianowski, J., Szala, L., Cegielska-Taras, T., Bartkowiak-Broda, I., & Gacek, K. (2020). Genetic Variation of Traits Affecting **Meal Quality** in eded Doubled Haploid Population of Winter Oilseed Rape. *Agronomy Research*, 18(3), 2259-2270. <https://doi.org/10.15159/ar.20.209>

- Maryan, K. E., Lahiji, H. S., Farrokhi, N., Haynes, P. A., Hamzelou, S., & Komeleh, H. H. (2020). Comparative Leaf Proteomics of *Brassica napus* Genotypes with Distinctive Levels of Early Cold Acclimation. Plant Molecular Biology Reporter, 1-18. <https://doi.org/10.1007/s11105-020-01249-4>
- Buhayov, V. D., & Vyshnevskyi, S. P. (2020). Influence of the winter rape hybrids rosette development on its productivity and winter resistance. Feeds and Feed Production, (89), 57-65. (Russian, English abstract) <https://doi.org/10.31073/kormovyrobnytstvo202089-05>
- Ma, L., Wu, J., Qi, W., Coulter, J. A., Fang, Y., Li, X., ... & Sun, W. (2020). Screening and verification of reference genes for analysis of gene expression in winter rapeseed (*Brassica rapa* L.) under abiotic stress. PloS one, 15(9), e0236577. <https://doi.org/10.1371/journal.pone.0236577>
- Eshkiki, E. M., Hajiahmadi, Z., Abedi, A., Kordrostami, M., & Jacquard, C. (2020). In Silico Analyses of Autophagy-Related Genes in Rapeseed (*Brassica napus* L.) under Different Abiotic Stresses and in Various Tissues. Plants, 9(10), 1393. <https://doi.org/10.3390/plants9101393>
- Gu, D., Zhou, X., Ma, Y., Xu, E., Yu, Y., Liu, Y., ... & Zhang, W. (2020). Expression of a *Brassica napus* metal transport protein (BnMTP3) in *Arabidopsis thaliana* confers tolerance to Zn and Mn. Plant Science, 110754. <https://doi.org/10.1016/j.plantsci.2020.110754>
- Shahzad, A.; Qian, M.; Sun, B.; Mahmood, U.; Li, S.; Fan, Y.; Chang, W.; Dai, L.; Zhu, H.; Li, J.; Qu, C.; Lu, K. Identification of Elite Rapeseed Drought-Tolerant Germplasm and Candidate Genes in a Natural Population of 265 Accessions. Preprints 2020, 2020090639 <https://www.preprints.org/manuscript/202009.0639/v1>
- Feng, Yn., Cui, Jq., Zhou, T. et al. Comprehensive dissection into morpho-physiologic responses, ionomic homeostasis, and transcriptomic profiling reveals the systematic resistance of allotetraploid rapeseed to salinity. BMC Plant Biol 20, 534 (2020). <https://doi.org/10.1186/s12870-020-02734-4>
- Yang, N., Li, S., Wang, S. et al. Dynamic transcriptome analysis indicates extensive and discrepant transcriptomic reprogramming of two rapeseed genotypes with contrasting NUE in response to nitrogen deficiency. Plant Soil 456, 369–390 (2020). <https://doi.org/10.1007/s11104-020-04720-z>
- Arifuzzaman, M., Rahman, M. Genome wide association mapping and candidate gene mining for root architectural traits in rapeseed/canola (*Brassica napus* L.) at late growth stage. Euphytica 216, 164 (2020). <https://doi.org/10.1007/s10681-020-02700-z>
- Shirokova, A. V., Volovik, V. T., Zagorskina, N. V., Zaitsev, G. P., Khudyakova, H. K., Korovina, L. M., ... & Baranova, E. N. (2020). From Dimness to Glossiness—Characteristics of the Spring Rapeseed Mutant Form without Glaucous Bloom (*Brassica napus* L.). Agronomy, 10(10), 1563. <https://doi.org/10.3390/agronomy10101563>
- Dai, C., Li, Y., Li, L. et al. An efficient Agrobacterium-mediated transformation method using hypocotyl as explants for *Brassica napus*. Mol Breeding 40, 96 (2020). <https://doi.org/10.1007/s11032-020-01174-0>

Islam, A. K. M. A., Era, F. M., & Chowdhury, N. K. (2020). Production of Restorer Lines from Segregating Progenies of *Brassica napus* L. Having Good Agronomic Value. International Journal of Applied Sciences and Biotechnology, 8(4), 400-409. <https://doi.org/10.3126/ijasbt.v8i4.31352>

## CROP PROTECTION

**Focus:** A publication which develops the ideas introduced by A. von Tiedemann in his keynote at the last IRC, in Berlin, June 2019. At that time, it provoked some reactions in the room: changing perspective is always difficult... but often necessary to build the future.

Zheng, X., Koopmann, B., Ulber, B., & von Tiedemann, A. (2020). A Global Survey on Diseases and Pests in Oilseed Rape—Current Challenges and Innovative Strategies of Control. Frontiers in Agronomy, 2, 1-15. <https://doi.org/10.3389/fagro.2020.590908>

KiTİŞ, Yasin Emre, GRENZ, Jan Hendrik, SAUERBORN, Joachim. "Effects of some cereal root exudates on germination of **broomrapes** (*Orobanche* spp. and *Phelipanche* spp.)". Mediterranean Agricultural Sciences 32 / 2 (August 2019): 145-150. <https://doi.org/10.29136/mediterranean.546564>

Bělonožníková, K., Vaverová, K., Vaněk, T., Kolařík, M., Hýsková, V., Vaňková, R., ... & Ryšlavá, H. (2020). Novel Insights into the Effect of **Pythium** Strains on Rapeseed Metabolism. Microorganisms, 8(10), 1472. <https://doi.org/10.3390/microorganisms8101472>

Rajvanshi, N. K., Singh, H. K., & Maurya, M. K. (2020). Management of **Alternaria blight** of Indian mustard through combo of seed treatment and foliar sprays of bioagent and fungicides. Journal of Pharmacognosy and Phytochemistry, 9(5), 1121-1123. <https://www.phytojournal.com/archives/2020/vol9issue5/PartP/9-5-187-409.pdf>

Munir, S., Shahzad, A. N., & Qureshi, M. K. (2020). Acuities into tolerance mechanisms via different bioassay during Brassicaceae-***Alternaria brassicicola*** interaction and its impact on yield. PloS one, 15(12), e0242545. <https://doi.org/10.1371/journal.pone.0242545>

Michael, P. J., Lui, K. Y., Thomson, L., Lamichhane, A., & Bennett, S. J. (2020). Impact of preconditioning temperature and duration period on carpogenic germination of diverse ***Sclerotinia sclerotiorum*** (Lib.) de Bary populations in south-western Australia. Plant Disease, (ja). <https://doi.org/10.1094/PDIS-09-20-1957-RE>

Starzycka-Korbas, E., Weber, Z., Matusczak, M. et al. The diversity of ***Sclerotinia sclerotiorum*** (Lib.) de Bary isolates from western Poland. J Plant Pathol (2020). <https://doi.org/10.1007/s42161-020-00705-0>

Dev, D., Tewari, A.K., Upadhyay, P. et al. Identification and nomenclature of ***Albugo candida*** pathotypes of Indian origin causing white rust disease of rapeseed-mustard. Eur J Plant Pathol 158, 987–1004 (2020). <https://doi.org/10.1007/s10658-020-02135-1>

Singh, O.W., Singh, N., Kamil, D. et al. Morpho-molecular variability and host reactivity of ***Albugo candida*** isolates infecting *Brassica juncea* genotypes in India. J Plant Pathol (2020). <https://doi.org/10.1007/s42161-020-00690-4>

- Murtza, T., You, M. P., & Barbetti, M. J. (2020). Canola Growth Stage at Time of Infection Determines Magnitude of **White Leaf Spot** (*Neopseudocercospora capsellae*) Impact. *Plant Disease*, (ja). <https://doi.org/10.1094/PDIS-09-20-2036-RE>
- Askarian Khanaman, H. (2020). Virulence and genetic structure of ***Plasmodiophora brassicae*** populations in Alberta, Canada. (PhD thesis). <https://era.library.ualberta.ca/items/4f415450-6b99-4b84-8245-d484f182e788>
- Hollman, K. B., Hwang, S. F., Manolii, V. P., & Strelkov, S. E. (2020). Pathotypes of ***Plasmodiophora brassicae*** collected from clubroot resistant canola (*Brassica napus L.*) cultivars in western Canada in 2017-2018. *Canadian Journal of Plant Pathology*. <https://doi.org/10.1080/07060661.2020.1851893>
- McDonald, M. R., Al-Daoud, F., Sedaghatkish, A., Moran, M., Cranmer, T. J., & Gossen, B. D. (2020). Changes in the range and virulence of ***Plasmodiophora brassicae*** across Canada. *Canadian Journal of Plant Pathology*, 1-7. <https://doi.org/10.1080/07060661.2020.1797882>
- Sedaghatkish, A., Gossen, B. D., & McDonald, M. R. (2020). Seed treatment of canola (*Brassica napus*) with the endomycorrhizal fungus *Piriformospora indica* does not reduce **clubroot**. *Canadian Journal of Plant Science*, (ja). <https://doi.org/10.1139/CJPS-2020-0126>
- Serdyuk, O., Trubina, V., & Gorlova, L. (2020). The evaluation of parental material of winter rapeseed (*Brassica napus L.*) and winter brown mustard (*Brassica juncea L.*) on resistance to **Phoma rot** in the central zone of the Krasnodar region of the Russian Federation. In E3S Web of Conferences (Vol. 222, p. 02030). EDP Sciences. (conference proceedings) <https://doi.org/10.1051/e3sconf/202022202030>
- Liu, F., Zou, Z., Huang, S. et al. Development of a specific marker for detection of a functional AvrLm9 allele and validating the interaction between AvrLm7 and AvrLm9 in ***Leptosphaeria maculans***. *Mol Biol Rep* 47, 7115–7123 (2020). <https://doi.org/10.1007/s11033-020-05779-8>
- Chen, Q., Peng, G., Kutcher, R., & Yu, F. (2020). Genome-wide DNA Variants Identify Genetic Diversity and Population Structure of ***Leptosphaeria maculans*** in Western Canada. (Preprint) <https://doi.org/10.21203/rs.3.rs-24766/v2>
- Rashid, M. H., Liban, S., Zhang, X., Parks, P., Borhan, H., & Fernando, W. D. (2021). Impact of *Brassica napus*–***Leptosphaeria maculans*** interaction on the emergence of virulent isolates of ***L. maculans***, causal agent of blackleg disease in canola. *Plant Pathology*, 70(2), 459-474. <https://doi.org/10.1111/ppa.13293>
- Liu, F., Zou, Z., Peng, G., & Fernando, W. D. (2020). ***Leptosphaeria maculans*** isolates reveal their allele frequency in western Canada. *Plant Disease*, (ja). <https://doi.org/10.1094/PDIS-08-20-1838-RE>
- Peng, G., Liu, C., Fernando, D. W., Lang, R., McLaren, D. L., Johnson, E. N., ... & Yu, F. (2020). Early fungicide treatment reduces **blackleg** on canola but yield benefit is realized only on susceptible cultivars under high disease pressure. *Canadian Journal of Plant Pathology*, 1-10. <https://doi.org/10.1080/07060661.2020.1824166>
- Soomro, W., Kutcher, R., Yu, F., Hwang, S. F., Fernando, D., Strelkov, S. E., & Peng, G. (2020). The race structure of ***Leptosphaeria maculans*** in western Canada between 2012 and 2014 and its influ-

- ence on blackleg of canola. Canadian Journal of Plant Pathology, 1-14. <https://doi.org/10.1080/07060661.2020.1829064>
- Cook, J., Douglas, G. M., Zhang, J., Glick, B. R., Langille, M. G., Liu, K. H., & Cheng, Z. (2020). Transcriptomic profiling of **Brassica napus** responses to **Pseudomonas aeruginosa**. Innate Immunity, 1753425920980512. <https://doi.org/10.1177/1753425920980512>
- Book chapter: Yadav S., Rathee M. (2020) **Sucking Pests** of Rapeseed-Mustard. In: Omkar (eds) Sucking Pests of Crops. Springer, Singapore. [https://doi.org/10.1007/978-981-15-6149-8\\_6](https://doi.org/10.1007/978-981-15-6149-8_6)
- Koirala, S. (2020). **Mustard Aphid** and Crop Production. International Journal of Applied Sciences and Biotechnology, 8(3), 310-317. <https://doi.org/10.3126/ijasbt.v8i3.31558>
- Mollaeei, M., Fathi, S. A. A., Nouri-Ganbalani, G., Hassanpour, M., & Golizadeh, A. (2020). Effects of strip intercropping of canola with faba bean, field pea, garlic, or wheat on control of **cabbage aphid** and crop yield. Plant Protection Science, 57(1), 59-65. <https://doi.org/10.17221/132/2019-PPS>
- Soni, S., & Kumar, S. (2020). Biological and behavioural characteristics of *Diaeretiella rapae* (McIntosh), a **parasitoid of Lipaphis erysimi** (Kaltenbach) infesting oilseed brassicas in India. Biocontrol Science and Technology, 1-18. <https://doi.org/10.1080/09583157.2020.1856331>
- Grocock, N. L., & Evenden, M. L. (2020). Local and Landscape-Scale Features Influence Bumble Bee (Hymenoptera: Apidae) Bycatch in **Bertha Armyworm** *Mamestra configurata* (Lepidoptera: Noctuidae) Pheromone-Baited Monitoring Traps. Environmental Entomology, 49(5), 1127-1136. <https://doi.org/10.1093/ee/nvaa087>
- Zhang, H., Xie, J., Fu, Y., Cheng, J., Qu, Z., Zhao, Z., ... & Jiang, D. (2020). A 2-kb Mycovirus Converts a Pathogenic Fungus into a Beneficial Endophyte for Brassica Protection and Yield Enhancement. Molecular Plant, 13(10), 1420-1433. <https://doi.org/10.1016/j.molp.2020.08.016>
- Qu, Z., Zhao, H., Zhang, H. et al. Bio-priming with a hypovirulent phytopathogenic fungus enhances the connection and strength of microbial interaction network in rapeseed. npj Biofilms Microbiomes 6, 45 (2020). <https://doi.org/10.1038/s41522-020-00157-5>
- EFSA (European Food Safety Authority), Anastassiadou, M., Bernasconi, G., Brancato, A., Carrasco Cabrera, L., Ferreira, L., ... & Verani, A. (2020). Modification of the existing maximum **residue levels for flupyradifurone** and DFA in rapeseeds/canola seeds and mustard seeds. EFSA Journal, 18(11), e06298. <https://doi.org/10.2903/j.efsa.2020.6298>
- Wood, S. C. (2020). Effects of Chronic **Neonicotinoid Exposure** on Saskatchewan Honey Bees (Doctoral dissertation, University of Saskatchewan). <https://harvest.usask.ca/handle/10388/13166>

## AGRONOMY

**Focus:** A special selection of German scientific publications proposed by Wolfgang Friedt, GCIRC President, and Dieter Bockey (UFOP), related to Rapeseed sustainability: cultivation, yield and nitrogen and greenhouse gas (Nitrous Oxide N<sub>2</sub>O) emissions:

Räbiger, T., Andres, M., Hegewald, H., Kesenheimer, K., Köbke, S., Quinones, T. S., ... & Kage, H. (2020). Indirect nitrous oxide emissions from oilseed rape cropping systems by NH<sub>3</sub> volatiliza-

- tion and nitrate leaching as affected by nitrogen source, N rate and site conditions. European Journal of Agronomy, 116, 126039. <https://doi.org/10.1016/j.eja.2020.126039>
- Kesenheimer, K., Pandeya, H. R., Müller, T., Buegger, F., & Ruser, R. (2019). Nitrous oxide emissions after incorporation of winter oilseed rape (*Brassica napus L.*) residues under two different tillage treatments. Journal of Plant Nutrition and Soil Science, 182(1), 48-59 <https://doi.org/10.1002/jpln.201700507>
- Ruser, R., Fuß, R., Andres, M., Hegewald, H., Kesenheimer, K., Köbke, S., ... & Flessa, H. (2017). Nitrous oxide emissions from winter oilseed rape cultivation. Agriculture, Ecosystems & Environment, 249, 57-69. <https://doi.org/10.1016/j.agee.2017.07.039>
- Köbke, S., Senbayram, M., Pfeiffer, B., Nacke, H., & Ditttert, K. (2018). Post-harvest N<sub>2</sub>O and CO<sub>2</sub> emissions related to plant residue incorporation of oilseed rape and barley straw depend on soil NO<sub>3</sub>-content. Soil and Tillage Research, 179, 105-113. <https://doi.org/10.1016/j.still.2018.01.013>
- Hegewald, H., Wensch-Dorendorf, M., Sieling, K., & Christen, O. (2018). Impacts of break crops and crop rotations on oilseed rape productivity: A review. European journal of agronomy, 101, 63-77. <https://doi.org/10.1016/j.eja.2018.08.003>
- Hegewald, H., Koblenz, B., Wensch-Dorendorf, M., & Christen, O. (2017). Yield, yield formation, and blackleg disease of oilseed rape cultivated in high-intensity crop rotations. Archives of Agronomy and Soil Science, 63(13), 1785-1799. <https://doi.org/10.1080/03650340.2017.1307508>
- Hegewald, H., Koblenz, B., Wensch-Dorendorf, M., & Christen, O. (2016). Impacts of high intensity crop rotation and N management on oilseed rape productivity in Germany. Crop and Pasture Science, 67(4), 439-449. <https://doi.org/10.1071/CP15214>
- Drastig, K., Quiñones, T. S., Zare, M., Dammer, K. H., & Prochnow, A. (2019). Rainfall interception by winter rapeseed in Brandenburg (Germany) under various nitrogen fertilization treatments. Agricultural and Forest Meteorology, 268, 308-317. <https://doi.org/10.1016/j.agrformet.2019.01.027>
- Pahlmann, I., Böttcher, U., Sieling, K., & Kage, H. (2013). Possible impact of the Renewable Energy Directive on N fertilization intensity and yield of winter oilseed rape in different cropping systems. Biomass and Bioenergy, 57, 168-179. <https://doi.org/10.1016/j.biombioe.2013.08.012>

Fieuza, R., Sicre, C. M., & Tallec, T. (2020). Towards an Improved Inventory of **N<sub>2</sub>O Emissions** Using Land Cover Maps Derived from Optical Remote Sensing Images. Atmosphere, 11(11), 1188. <https://doi.org/10.3390/atmos1111118>

Flénet, F., Wagner, D., & Simonin, P. (2020). Examination of an attempt to improve rapeseed cultivation in France in order to reduce the **greenhouse gas emissions** of biodiesel. OCL, 27, 69. <https://doi.org/10.1051/ocl/2020068>

Glenn, A. J., Moulin, A. P., Roy, A. K., & Wilson, H. F. (2021). Soil **nitrous oxide emissions** from no-till canola production under variable rate nitrogen fertilizer management. Geoderma, 385, 114857. <https://doi.org/10.1016/j.geoderma.2020.114857>

- Moradi Aghdam A., Sayfzadeh S., Shirani Rad A.H., Valadabadi S.A., Zakerin H.R. The assessment of **water stress and delay cropping** on quantitative and qualitative traits of rapeseed genotypes. Industrial Crops and Products, Volume 131, 2019, <https://doi.org/10.1016/j.indcrop.2019.01.051>
- Rahimi-Moghaddam, S., Eyni-Nargese, H., Ahmadi, S. A. K., & Azizi, K. Towards withholding **irrigation regimes and drought-resistant genotypes** as strategies to increase canola production in drought-prone environments: A modeling approach. Agricultural Water Management, 243, 106487. <https://doi.org/10.1016/j.agwat.2020.106487>
- Shafiqhi, A., Ardakani, M. R., Rad, A. H. S., Alavifazel, M., & Rafiei, F. (2020). Grain yield and associated physiological traits of rapeseed (*Brassica napus L.*) cultivars under different planting dates and drought stress at the 3 flowering stage. Italian Journal of Agronomy. <https://doi.org/10.4081/ija.2020.1648>
- Feizabadi, A., Noormohammadi, G. & Fatehi, F. Changes in Growth, Physiology, and Fatty Acid Profile of Rapeseed Cultivars Treated with Vermicompost Under **Drought Stress**. J Soil Sci Plant Nutr (2020). <https://doi.org/10.1007/s42729-020-00353-4>
- Stassinos, P. M., Rossi, M., Borromeo, I., Capo, C., Beninati, S., & Forni, C. (2020). Amelioration of **salt stress tolerance** in rapeseed (*Brassica napus*) cultivars by seed inoculation with *Arthrobacter globiformis*. Plant Biosystems-An International Journal Dealing with all Aspects of Plant Biology, 1-12. <https://doi.org/10.1080/11263504.2020.1857872>
- Secchi, M. A., Bastos, L. M., Stamm, M. J., Wright, Y., Foster, C., Messina, C. D., & Ciampitti, I. A. (2021). **Winter survival** response of canola to meteorological variables and adaptative areas for current canola germplasm in the United States. Agricultural and Forest Meteorology, 297, 108267. <https://doi.org/10.1016/j.agrformet.2020.108267>
- Riar, A., Gill, G., & McDonald, G. K. (2020). Rate of Nitrogen Rather Than Timing of Application Influence Yield and **NUE** of Canola in South Australian Mediterranean Environments. Agronomy, 10(10), 1505. <https://doi.org/10.3390/agronomy10101505>
- D. Condaminet, A. Zimmermann, B. Billiot, B. Crémilleux and S. Pluchon, "Using **Data Science** to Improve the Identification of Plant **Nutritional Status**," 2020 IEEE 7th International Conference on Data Science and Advanced Analytics (DSAA), sydney, Australia, 2020, pp. 496-505, <https://doi.org/10.1109/DSAA49011.2020.00064>
- Ma, B.L., Zheng, Z.M., de Silva, N. et al. Graphical analysis of nitrogen and **sulfur supply** on yield and related traits of canola in eastern Canada. Nutr Cycl Agroecosyst 118, 293–309 (2020). <https://doi.org/10.1007/s10705-020-10097-3>
- Rosa, W. B., Júnior, J. B. D., Tomm, G. O., da Costa, A. C. T., & Queiroz, S. B. (2020). Optimization of canola agronomic yield submitted to different doses of **potassium in flowering**. Revista Engenharia na agricultura Reveng 28, 389-396. <https://doi.org/10.13083/reveng.v29i1.8201>
- Amina, I., & Abdelhalim, A. K. Impact of potassium fertilization on growth, yield and water productivity of canola under water stress condition. [http://plantarchives.org/20-2/8389-8402%20\(6958\).pdf](http://plantarchives.org/20-2/8389-8402%20(6958).pdf)

- Tian, C., Zhou, X., Liu, Q., Peng, J., Zhang, Z., Song, H., ... & Abou-Elwafa, S. F. (2020). Increasing yield, quality and profitability of winter oilseed rape (*Brassica napus*) under combinations of nutrient levels in fertiliser and planting density. *Crop and Pasture Science*, 71(12), 1010-1019. <https://doi.org/10.1071/CP20328>
- Hartman, M. D., & Jeffrey, S. R. (2020). Estimating the economic optimal target **density of hybrid canola** based on data from a western Canadian meta-analysis. *Canadian Journal of Plant Science*, (ja). <https://doi.org/10.1139/CJPS-2020-0162>
- Korzeniowska, J., Stanisławska-Glubiak, E., & Lipiński, W. (2020). New limit values of **micronutrient deficiency** in soil determined using 1 M HCl extractant for wheat and rapeseed. *SOIL SCIENCE ANNUAL*, 71(3), 205-214. <https://doi.org/10.37501/soilsa/126079>
- Khan, Z., Zhang, K., Khan, M. N., Fahad, S., Xu, Z., & Hu, L. (2020). Coupling of **Biochar** with Nitrogen Supplements Improve **Soil Fertility**, Nitrogen Utilization Efficiency and Rapeseed Growth. *Agronomy*, 10(11), 1661. <https://doi.org/10.3390/agronomy10111661>
- Jiao, Y., Acdan, J., Xu, R., Deventer, M. J., Zhang, W., & Rhew, R. C. (2020). Global **methyl halide emissions** from rapeseed (*Brassica napus*) using life cycle measurements. *Geophysical Research Letters*, 47(19), e2020GL089373. <https://doi.org/10.1029/2020GL089373>
- Dowling, A., Sadras, V. O., Roberts, P., Doolette, A., Zhou, Y., & Denton, M. D. **Legume-oilseed intercropping** in mechanised broadacre agriculture—a review. *Field Crops Research*, 260, 107980. <https://doi.org/10.1016/j.fcr.2020.107980>
- Laishram, R. C., Sorokhaibam, S., Leitam, O. C., Wangkheirakpam, M., & Narendrajit, C. H. Yield, economics and biological indices of **chickpea+ rapeseed intercropping** system as influenced by moisture conservation and nutrient management practices under rainfed conditions. [REFERENCE](#)
- Charles, R., Baux, A., Dierauer, H., & Daniel, C. (2020). **Organic rapeseed in Switzerland**: 20 years of practice. *OCL*, 27, 68. <https://doi.org/10.1051/ocl/2020055>
- Zuo, Q., Wang, L., Zheng, J., You, J., Yang, G., Leng, S., & Liu, J. (2020). Effects of uniconazole rate on agronomic traits and physiological indexes of rapeseed blanket seedling. *Oil Crop Science*. <https://doi.org/10.1016/j.ocsci.2020.12.003>
- KC, K.B., Montocchio, D., Berg, A. et al. **How climatic and sociotechnical factors influence** crop production: a case study of canola production. *SN Appl. Sci.* 2, 2063 (2020). <https://doi.org/10.1007/s42452-020-03824-6>
- Loveimi, N., Akram, A., Bagheri, N., & Hajiahmad, A. (2021). Evaluation of Several Spectral Indices for **Estimation of Canola Yield** using **Sentinel-2** Sensor Images. *Journal of Agricultural Machinery*, 11(2), 447-464. <https://doi.org/10.22067/jam.v1i2.80232>
- Jiménez-Gómez, A., Saati-Santamaría, Z., Kostovcik, M., Rivas, R., Velázquez, E., Mateos, P. F., ... & García-Fraile, P. (2020). Selection of the Root Endophyte *Pseudomonas brassicacearum* CDVBN10 as **Plant Growth Promoter** for *Brassica napus* L. *Crops. Agronomy*, 10(11), 1788. <https://doi.org/10.3390/agronomy10111788>
- Zh, B. R., Mukasheva, T. D., Sydykbekova, R. K., Ignatova, L. V., Omirbekova, A. A., Davenova, N. A., & Esentaeva, K. (2020). **Plant growth-promoting bacteria** isolated from agrocenoses of agricul-

- tural plants. Eurasian Journal of Ecology, 65(4), 17-24. <https://doi.org/10.26577/EJE.2020.v65.i4.02>.
- Premachandra, D., Hudek, L., Enez, A., Ballard, R., Barnett, S., Franco, C. M., & Brau, L. (2020). Assessment of the Capacity of **Beneficial Bacterial Inoculants** to Enhance Canola (*Brassica napus L.*) Growth under Low Water Activity. Agronomy, 10(9), 1449. <https://doi.org/10.3390/agronomy10091449>
- El-Shazly, M. M. Effect of using **mycorrhizae and biostimulants** on productivity of canola under salt stress. [http://plantarchives.org/20-2/8303-8314%20\(7094\).pdf](http://plantarchives.org/20-2/8303-8314%20(7094).pdf)
- Suchan, D. M., Bergsveinson, J., Manzon, L., Pierce, A., Kryachko, Y., Korber, D., ... & Yost, C. K. (2020). Transcriptomics reveal core activities of the **plant growth-promoting bacterium *Delftia acidovorans* RAY209** during interaction with canola and soybean roots. Microbial genomics, 6(11). <https://doi.org/10.1099/mgen.0.000462>
- Rezgui, C., Riah-Anglet, W., Benoit, M., Bernard, P. Y., Laval, K., & Trinsoutrot-Gattin, I. (2020). Impacts of the Winter Pea Crop (Instead of Rapeseed) on **Soil Microbial Communities**, Nitrogen Balance and Wheat Yield. Agriculture, 10(11), 548. <https://doi.org/10.3390/agriculture10110548>
- Aly, M. M., Shehata, H. S., Elareny, I. M., Khalifa, D. M., & El-Tapey, H. M. A. **Rhizo-microbiota** are among the pillars for proper canola (*Brassica napus L.*) and sesame (*Sesamum indicum L.*) production in marginal soil. <http://www.curesweb.com/mejas/mejas/2020/mejas.2020.10.4.64.pdf>
- Lupwayi, N. Z., Schwinghamer, T. D., Tidemann, B. D., Kubota, H., Turkington, T. K., Khakbazan, M., & St. Luce, M. Causal relationships from **legume crops to soil microbial properties** relative to canola. Agronomy Journal. <https://doi.org/10.1002/agi2.20493>
- Hirzel, J., Undurraga, P., León, L., Panichini, M., Carrasco, J., González, J., & Matus, I. (2020). Canola production and effect on **soil chemical properties** in response to different residue levels from three biannual **crop rotations**. Plant Production Science, 1-10. <https://doi.org/10.1080/1343943X.2020.1851142>
- Mazzilli, S. R., Abbate, S., Silva, H., & Mendoza, Y. (2020). ***Apis mellifera*** visitation enhances productivity in rapeseed. Journal of Apicultural Research, 1-9. <https://doi.org/10.1080/00218839.2020.1856558>
- Pilla, T. P., Galon, L., Brandler, D., Bianchessi, F., Tonin, R. J., Toni, J. R., ... & Vargas, L. (2020). Physical, Physiological and Sanitary Quality of Canola Seeds After the Application of **Maturing Herbicides**. Journal of Agricultural Studies, 8(3), 721-736. <http://www.macrothink.org/journal/index.php/jas/article/download/16756/13256>
- Schaafsma, A. W., & Limay-Rios, V. (2020). Fugitive Dust During Planting of Canola with an Air Seeder as a Source of **Environmental Contamination for Pesticides** Applied on **Seed**: A Case Study. Environmental Toxicology and Chemistry, 39(12), 2420-2423. <https://doi.org/10.1002/etc.4892>

## PHYSIOLOGY

**Focus:** Kirkegaard JA, Lilley JM, Berry PM, Rondanini DP (2020) **Canola**. Book chapter in “Crop Physiology Case Histories for Major Crops” V. Sadras and D. Calderini. (Eds.) Academic Press ISBN: 9780128191941.

- Rys, M., Pociecha, E., Oliwa, J., Ostrowska, A., Jurczyk, B., Saja, D., & Janeczko, A. (2020). **Deacclimation of Winter Oilseed Rape—Insight into Physiological Changes**. *Agronomy*, 10(10), 1565. <https://doi.org/10.3390/agronomy10101565>
- Sadras and D. Calderini. (Eds.) Academic Press ISBN: 9780128191941.
- Zaheer, I. E., Ali, S., Saleem, M. H., Arslan Ashraf, M., Ali, Q., Abbas, Z., ... & Wijaya, L. (2020). Zinc-lysine supplementation mitigates oxidative stress in rapeseed (*Brassica napus L.*) by preventing **phytotoxicity of chromium**, when irrigated with tannery wastewater. *Plants*, 9(9), 1145. <https://doi.org/10.3390/plants9091145>
- Raza, A. Eco-physiological and Biochemical Responses of Rapeseed (*Brassica napus L.*) to **Abiotic Stresses**: Consequences and Mitigation Strategies. *J Plant Growth Regul* (2020). <https://doi.org/10.1007/s00344-020-10231-z>
- Hu, J., Zhang, F., Gao, G., Li, H., & Wu, X. Auxin-related genes associated with leaf petiole angle at the seedling stage are involved in **adaptation to low temperature** in *Brassica napus*. *Environmental and Experimental Botany*, 182, 104308. <https://doi.org/10.1016/j.envexpbot.2020.104308>
- He, X. (2020). An Insight into the Responses of Early-Maturing *Brassica napus* to Different **Low-Temperature Stresses**. *Abiotic Stress in Plants*. <https://doi.org/10.5772/intechopen.93708>
- Whish, J. P. M., Lilley, J. M., Morrison, M. J., Cocks, B., & Bullock, M. (2020). **Vernalisation** in Australian spring canola explains variable flowering responses. *Field Crops Research*, 258, 107968. <https://doi.org/10.1016/j.fcr.2020.107968>
- Chen, S., Stefanova, K., Siddique, K. H., & Cowling, W. A. (2020). Transient daily **heat stress** during the early reproductive phase disrupts **pod and seed development** in *Brassica napus L.* *Food and Energy Security*, e262. <https://doi.org/10.1002/fes3.262>
- Wu, W., Duncan, R. W., & Ma, B. L. (2021). The stage sensitivity of short-term **heat stress** to lodging-resistant traits and yield determination in canola (*Brassica napus L.*). *Journal of Agronomy and Crop Science*, 207(1), 74-87. <https://doi.org/10.1111/jac.12464>
- Zhang, W., Cao, H., Zhang, W., Hanan, J. S., Ge, D., Cao, J., ... & Wu, F. (2020). An aboveground **biomass partitioning** coefficient model for rapeseed (*Brassica napus L.*). *Field Crops Research*, 259, 107966. <https://doi.org/10.1016/j.fcr.2020.107966>
- Adavi, Z., Sadat Asilan, K., & Baghbani-Arani, A. (2020). Effect of **Paclobutrazol on Canopy Temperature** and Some Quantitative and Qualitative Characteristics of Two Rapeseed (*Brassica napus L.*) Cultivars in Different Irrigation Regimes. *Isfahan University of Technology-Journal of Crop Production and Processing*, 10(3), 73-88. (Persian, English abstract) [https://jcpp.iut.ac.ir/browse.php?a\\_id=2969&sid=1&slc\\_lang=en](https://jcpp.iut.ac.ir/browse.php?a_id=2969&sid=1&slc_lang=en)

- Gorzin, M., Ghaderi-Far, F., Sadeghipour, H.R. et al. **Induced Thermo-dormancy** in Rapeseed (*Brassica napus L.*) Cultivars by Sub- and Supra-optimal Temperatures. *J Plant Growth Regul* (2020). <https://doi.org/10.1007/s00344-020-10266-2>
- Molnár, K., Biró-Janka, B., Nyárádi, I. I., Fodorpataki, L., Varga, B. E., Bálint, J., & Duda, M. M. (2020). Effects of **Priming** with Ascorbic Acid, L-Cysteine and Triacontanol on Germination of Rapeseed (*Brassica napus L.*). *Acta Biologica Marisiensis*, 3(2), 48-55. <https://doi.org/10.2478/abmj-2020-0010>
- Feng, X., An, Y., Gao, J., & Wang, L. (2020). Photosynthetic Responses of Canola to Exogenous Application or Endogenous Overproduction of 5-Aminolevulinic Acid (ALA) under Various Nitrogen Levels. *Plants*, 9(11), 1419. <https://doi.org/10.3390/plants9111419>
- Martel, A. B., Taylor, A. E., & Qaderi, M. M. (2020). Individual and interactive effects of **temperature and light intensity** on canola growth, physiological characteristics and methane emissions. *Plant Physiology and Biochemistry*, 157, 160-168. <https://doi.org/10.1016/j.plaphy.2020.10.016>
- BISWAS, N., YADAV, S., YADAV, S., CHOUDHARY, R., SAINI, N., DAHUJA, A., ... & YADAV, D. K. (2020). **Vigor** difference during storage and germination in Indian mustard explained by reactive oxygen species and antioxidant enzymes. *Turkish Journal of Agriculture and Forestry*, 44(6), 577-588. <https://journals.tubitak.gov.tr/agriculture/abstract.htm?id=28264>

### ***PROCESSING and USES***

- Rokosik, Ewa, Krzysztof Dwiecki, and Aleksander Siger. "The quality of **cold-pressed rapeseed oil** obtained from seeds of *Brassica napus L.* with increased moisture content." *Acta Scientiarum Polonorum Technologia Alimentaria* 18.2 (2019): 205-218. <https://doi.org/10.17306/J.AFS.2019.0672>
- Kmiecik, D., Fedko, M., Rudzińska, M., Siger, A., Gramza-Michałowska, A., & Kobus-Cisowska, J. (2021). **Thermo-Oxidation of Phytosterol Molecules** in Rapeseed Oil during Heating: The Impact of Unsaturation Level of the Oil. *Foods*, 10(1), 50. <https://doi.org/10.3390/foods10010050>
- Gaber, M. A. F. M., Juliano, P., Mansour, M. P., Shrestha, P., Taylor, C., Smith, R., & Trujillo, F. J. (2020). Improvement of the Canola Oil **Degumming Process** by Applying a Megasonic Treatment. *Industrial Crops and Products*, 158, 112992. <https://doi.org/10.1016/j.indcrop.2020.112992>
- Kruse, M., Kemper, M., Gancheva, S., Osterhoff, M., Dannenberger, D., Markgraf, D., ... & Pfeiffer, A. F. (2020). **Dietary Rapeseed Oil Supplementation** Reduces Hepatic Steatosis in Obese Men—A Randomized Controlled Trial. *Molecular Nutrition & Food Research*, 64(21), 2000419. <https://doi.org/10.1002/mnfr.202000419>
- Robert, C., Couëdelo, L., Knibbe, C., Fonseca, L., Buisson, C., Errazuriz-Cerda, E., ... & Michalski, M. C. (2020). **Rapeseed Lecithin** Increases Lymphatic Lipid Output and α-Linolenic Acid Bioavailability in Rats. *The Journal of Nutrition*, 150(11), 2900-2911. <https://doi.org/10.1093/jn/nxaa244>
- David, A., David, M., Lesniarek, P., Corfias, E., Pululu, Y., Delample, M., & Snabre, P. (2020). **Oleogellation** of rapeseed oil with cellulose fibers as an innovative strategy for **palm oil substitution** in

chocolate spreads. Journal of Food Engineering, 292, 110315.  
<https://doi.org/10.1016/j.jfoodeng.2020.110315>

Ettl, J., Bernhardt, H., Huber, G. et al. Evaluation of pure **rapeseed oil as a renewable fuel** for agricultural machinery based on emission characteristics and long-term operation behaviour of a fleet of 18 tractors. SN Appl. Sci. 2, 1711 (2020). <https://doi.org/10.1007/s42452-020-03490-8>

Emberger, P., Hinrichs, M., Huber, G., Emberger-Klein, A., Thuneke, K., Pickel, P., & Remmele, E. Field tests and real-world exhaust gas emissions of a pure **rapeseed oil-fuelled** harvester in forestry: Testing a solution for combined water, soil, and climate protection. Journal of Cleaner Production, 280, 124360. <https://doi.org/10.1016/j.jclepro.2020.124360>

Abelniece, Z., & Kampars, V. (2020). Studying **the kinetics** of rapeseed oil reactions with methanol, methyl formate, and methyl acetate under mild conditions for **biodiesel** production. Biofuels, 1-8. <https://doi.org/10.1080/17597269.2020.1827929>

D. Kurczyński, "Impact of **RME biodiesel** on the concentration of **particulates and nitrogen oxides** in compression ignition engine exhaust fumes," 2020 XII International Science-Technical Conference AUTOMOTIVE SAFETY, Kielce, 2020, pp. 1-7, <https://doi.org/10.1109/AUTOMOTIVESAFETY47494.2020.9293517>

Vithya P., Sriram G., Arumugam S. (2021) RSM-Based Optimization of Process Parameters in Synthesis of **Pentaerythritol Ester of Rapeseed Oil**. In: Rajmohan T., Palanikumar K., Davim J.P. (eds) Advances in Materials and Manufacturing Engineering. Springer Proceedings in Materials, vol 7. Springer, Singapore. [https://doi.org/10.1007/978-981-15-6267-9\\_58](https://doi.org/10.1007/978-981-15-6267-9_58)

Uram, K., Prociak, A., & Kurańska, M. (2020). Influence of the chemical structure of rapeseed oil-based polyols on selected properties of **polyurethane foams**. Polimery, 65(10), 698-707. <https://doi.org/10.14314/polimery.2020.10.5>

Gu, K., Lin, W., Yuan, X., Peng, H., Wang, S., Lv, J., & Zhu, Z. (2020). Tribological performance and mechanism of 2D calcium borate nanoslice capped with stearic acid in rapeseed oil. Journal of Dispersion Science and Technology, 1-11. <https://doi.org/10.1080/01932691.2020.1844012>

Majcher, J. (2020). The use of an electroseparator with bifilar winding for **extracting germ** from crushed rape seed. Przegląd Elektrotechniczny, 96. <https://doi.org/10.15199/48.2020.09.15> or [REFERENCE](#)

WYRICK, Meghan K., XIA, Min, et SLESSAREVA, Janna. Method and means for an **isolation of membrane-bound proteins** from a biological sample, preferably processed plant seed meal. U.S. Patent Application No 16/956,058, 26 nov. 2020. <https://patents.google.com/patent/US20200369717A1/en>

Fetzer, A., Müller, K., Schmid, M., & Eisner, P. (2020). **Rapeseed proteins for technical applications:** Processing, isolation, modification and functional properties—A **review**. Industrial Crops and Products, 158, 112986. <https://doi.org/10.1016/j.indcrop.2020.112986>

Chmielewska, A., Kozłowska, M., Rachwał, D., Wnukowski, P., Amarowicz, R., Nebesny, E., & Rosicka-Kaczmarek, J. (2020). **Canola/rapeseed protein**—nutritional value, functionality and food application: a **review**. Critical Reviews in Food Science and Nutrition, 1-21. <https://doi.org/10.1080/10408398.2020.1809342>

- Kotecka-Majchrzak, K., Sumara, A., Fornal, E., & Montowska, M. (2020). **Oilseed proteins—properties and application as a food ingredient.** Trends in Food Science & Technology. <https://doi.org/10.1016/j.tifs.2020.10.004>
- Li, X., Shi, J., Scanlon, M., Xue, S. J., & Lu, J. (2021). Effects of **pretreatments** on physicochemical and structural properties of **proteins** isolated from canola seeds after oil extraction by **supercritical-CO<sub>2</sub> process**. LWT, 137, 110415. <https://doi.org/10.1016/j.lwt.2020.110415>
- Ntöne, E., van Wesel, T., Sagis, L. M., Meinders, M., Bitter, J. H., & Nikiforidis, C. V. (2021). Adsorption of **rapeseed proteins** at oil/water interfaces. Janus-like napins dominate the interface. Journal of Colloid and Interface Science, 583, 459-469. <https://doi.org/10.1016/j.jcis.2020.09.039> or <https://edepot.wur.nl/532719>
- Banovic, M., & Sveinsdóttir, K. (2020). Importance of Being Analogue: Female Attitudes towards **Meat Analogue containing Rapeseed Protein**. Food Control, 107833. <https://doi.org/10.1016/j.foodcont.2020.107833>
- Bailey, H. M., & Stein, H. H. (2020). PSIX-2 **Amino acid digestibility** and digestible indispensable amino acid scores of a few protein isolates and concentrates derived from cereal grains, plant proteins, or dairy proteins. Journal of Animal Science, 98(Supplement\_3), 186-187. <https://doi.org/10.1093/jas/skaa054.329>
- Tang, Y. R., & Ghosh, S. (2021). Stability and rheology of **canola protein isolate**-stabilized concentrated oil-in-water **emulsions**. Food Hydrocolloids, 113, 106399. <https://doi.org/10.1016/j.foodhyd.2020.106399>
- US Patent: DSM IP ASSETS B.V., Heerlen (NL) / SHI, Jing. **Foam comprising rapeseed and dairy proteins.** U.S. Patent Application No 16/959,768, 19 nov. 2020. <https://patents.google.com/patent/US20200359650A1/en>
- Barzegar, M., Behrooz, R., Mansouri, H. R., Najafi, S. K., Lorenz, L. F., & Frihart, C. R. (2020). Comparison of canola and soy flour with added isocyanate as **wood adhesives**. Journal of the American Oil Chemists' Society. <https://doi.org/10.1002/aocs.12410>
- Dražbo, A. A., Juśkiewicz, J., Józefiak, A., & Konieczka, P. (2020). The **Fermentation** Process Improves the Nutritional Value of **Rapeseed Cake** for Turkeys—Effects on Performance, Gut Bacterial Population and Its Fermentative Activity. Animals, 10(9), 1711. <https://doi.org/10.3390/ani10091711>
- Zhang, X., Wang, H., Zhang, J., Lin, B., Chen, L., Wang, Q., ... & Deng, J. (2020). Assessment of rapeseed meal as **fish meal alternative** in diets for juvenile Asian red-tailed catfish (*Hemibagrus wyckiooides*). Aquaculture Reports, 18, 100497. <https://doi.org/10.1016/j.agrep.2020.100497>
- Hernández, C., Olmeda-Guerrero, L., Chávez-Sánchez, M. C., Ibarra-Castro, L., Gaxiola-Cortez, G., & Martínez-Cárdenas, L. (2020). Nutritional evaluation of canola meal as **fish meal** replacement for juvenile spotted rose snapper (*Lutjanus guttatus*): Effects on growth performance, hematological parameters, body composition, and nutrient digestibility. Animal Feed Science and Technology, 269, 114683. <https://doi.org/10.1016/j.anifeedsci.2020.114683>

- Li, P., Lyu, Z., Wang, L., Huang, B., & Lai, C. (2020). **Nutritive values** of double-low rapeseed expellers and rapeseed meal with or without supplementation of multi-enzyme in **pigs**. Canadian Journal of Animal Science, 100(4), 729-738. <https://doi.org/10.1139/cjas-2019-0097>
- Lee, J. W., Kim, I. H., & Woyengo, T. A. (2020). Toxicity of Canola-Derived **Glucosinolate** Degradation Products in **Pigs**—A Review. Animals, 10(12), 2337. <https://doi.org/10.3390/ani10122337>
- Bujňák, L., Nadš, P., Skalická, M., & Marcinčák, S. (2020). Effects of Long-Term Feeding of **Treated Rapeseed Meal** on Growth Performance, Blood Mineral Profile and Fatty Acid Composition of Back Fat in **Pigs**. Folia Veterinaria, 64(4), 37-43. <https://doi.org/10.2478/fv-2020-0035>
- Gao, M., Cieślak, A., Kierończyk, B., Huang, H., Yanza, Y. R., Zaworska-Zakrzewska, A., ... & Szumacher-Strabel, M. (2020). Effects of **Raw and Fermented Rapeseed Cake** on Growth Performance, Methane Production, and Breast Meat Fatty Acid Composition in **Broiler Chickens**. Animals, 10(12), 2250. <https://doi.org/10.3390/ani10122250>
- Heim, R., & Krebs, G. (2020). Utilisation of **canola meal as protein source in dairy cow** diets: a review. Agriculture and Natural Resources, 54(6), 623-632. <https://li01.tci-thaijo.org/index.php/anres/article/view/248673/170044>
- Samková, E., & Kalač, P. (2020). Rapeseed supplements affect propitiously fatty acid **composition of cow milk fat**: A meta-analysis. Livestock Science, 104382. <https://doi.org/10.1016/j.livsci.2020.104382>
- Białek, M., Czuderna, M., & Zaworski, K. (2020). Diets enriched in fish and **rapeseed oils**, carnosic acid, and different chemical forms of selenium affect fatty acid profile in the periintestinal fat and indices of nutritional properties of selected tissues of **lambs**. Agricultural and Food Science, 29(5), 405-419. <https://doi.org/10.23986/afsci.97267>
- Sekali, M., Mlambo, V., Marume, U., & Mathuthu, M. (2020). Replacement of Soybean Meal with **Heat-Treated Canola Meal** in Finishing Diets of Meatmaster **Lambs**: Physiological and Meat Quality Responses. Animals, 10(10), 1735. <https://doi.org/10.3390/ani10101735>
- Laguna, O., Guyot, S., Yu, X., Broudiscou, L. P., Chapoutot, P., Solé-Jamault, V., ... & Dauguet, S. (2020). The PHENOLO project or how to separate and add-value to **phenolic compounds** present in rapeseed and sunflower **meals**. OCL Oilseeds and fats crops and lipids, 27, 61. <https://doi.org/10.1051/ocl/2020056>
- Nandasiri, R., Eskin, N. M., Komatsu, E., Perreault, H., & Thiayam-Holländer, U. (2020). Valorization of canola by-products: Concomitance of **flavor-active bitter phenolics** using pressurized heat treatments. LWT, 110397. <https://doi.org/10.1016/j.lwt.2020.110397>
- Chai, X., Xu, L., Li, Y., Qiu, J., Li, Y., Lv, L., & Zhu, Y. (2020). Development and Experimental Analysis of a Fuzzy Grey Control System on Rapeseed **Cleaning Loss**. Electronics, 9(11), 1764. <https://doi.org/10.3390/electronics9111764>
- Wawrzyniak, J. (2020). Application of Artificial Neural Networks to Assess the **Mycological State of Bulk Stored Rapeseeds**. Agriculture, 10(11), 567. <https://doi.org/10.3390/agriculture10110567>
- Jia, X., Wang, L., Zheng, C., Yang, Y., Wang, X., Hui, J., & Zhou, Q. (2020). Key **Odorant Differences** in Fragrant *Brassica napus* and *Brassica juncea* **Oils** Revealed by Gas Chromatography–

Olfactometry, Odor Activity Values, and Aroma Recombination. Journal of Agricultural and Food Chemistry, 68(50), 14950-14960. <https://doi.org/10.1021/acs.jafc.0c05944>

Demeke, T., Eng, M., Holigroski, M. et al. Effect of Amount of DNA on Digital PCR Assessment of Genetically Engineered Canola and Soybean Events. Food Anal. Methods 14, 372–379 (2021). <https://doi.org/10.1007/s12161-020-01889-y>

Veluri, S., & Olukosi, O. A. (2020). Metabolizable Energy of Soybean Meal and Canola Meal as Influenced by the Reference Diet Used and Assay Method. Animals, 10(11), 2132. <https://doi.org/10.3390/ani10112132>

## ECONOMY and MARKET

Deepayan Debnath, Jarrett Whistance, Patrick Westhoff, Mike Helmar, Chapter 9 - Consequences of US and EU biodiesel policies on global food security. Editor(s): Deepayan Debnath, Suresh Chandra Babu. Biofuels, Bioenergy and Food Security, Academic Press, 2019, <https://doi.org/10.1016/B978-0-12-803954-0.00009-7>

Tian, Z., Ji, Y., Xu, H., Qiu, H., Sun, L., Zhong, H., & Liu, J. (2020). The potential contribution of growing rapeseed in winter fallow fields across Yangtze River Basin to energy and food security in China. Resources, Conservation and Recycling, 164, 105159. <https://doi.org/10.1016/j.resconrec.2020.105159>

Chauhan, J., Choudhury, P., Pal, S., & Singh, K. Analysis of seed chain and its implication in rapeseed-mustard (*Brassica spp.*) production in India. THE INDIAN SOCIETY OF OILSEEDS RESEARCH, 71. [REFERENCE](#)

Wells, J., & Slade, P. (2020). The effect of the Canada–China canola trade dispute on canola prices. Canadian Journal of Agricultural Economics/Revue canadienne d'agroéconomie. <https://doi.org/10.1111/cjag.12258>

Sun, S. (2020). China's Ban on Canadian Canola: Reasons, Impacts, and Policy Perspectives. <https://doi.org/10.7939/r3-bzhn-d142>

Gularte, J. A., Macedo, V. G. K., & Panizzo, L. E. (2020). Canola seed production and market in Brazil. Applied Research & Agrotechnology, 13, 5834-1. (Portuguese, English abstract) <https://revistas.unicentro.br/index.php/repaa/article/view/5834>

Santeramo, F. G., Di Gioia, L., & Lamonaca, E. (2020). Price responsiveness of supply and acreage in the EU vegetable oil markets: policy implications. Land Use Policy, 105102. <https://doi.org/10.1016/j.landusepol.2020.105102> or [REFERENCE](#)

Azam, A. H. M., Sarmidi, T., Md Nor, A. H. S., & Zainuddin, M. R. K. (2020). Co-movement among world vegetable oil prices: a wavelet-based analysis. International Journal of Business & Society, 21(3). <http://www.ijbs.unimas.my/images/repository/pdf/Vol21-no3-paper6.pdf>

## **MUSTARD and Other Brassicae**

Karpavičienė, Birutė, Nijolė Maršalkienė, and Liuda Žilénaitė. "Seed composition of different **Came-lina sativa** and **Crambe abyssinica** cultivars." 26th NJF Congress: Agriculture for the Next 100 Years, 27-29 June, 2018 Kaunas r. Lithuania: Programme and Summaries of Presentations. Akademija, 2018. 2018. <https://hdl.handle.net/20.500.12259/92459>

Samarappuli, D., Zanetti, F., Berzuini, S., & Berti, M. T. (2020). **Crambe** (*Crambe abyssinica* Hochst): A non-food oilseed crop with great potential: A review. *Agronomy*, 10(9), 1380. <https://doi.org/10.3390/agronomy10091380>

Mohdaly, A. A. A., & Ramadan, M. F. (2020). Characteristics, composition and functional properties of seeds, seed cake and seed oil from different **Brassica carinata** genotypes. *Food Bioscience*, 100752. <https://doi.org/10.1016/j.fbio.2020.100752>

Pal, S., & Debnath, P. Seasonal incidence of major **insect pests and predators** of mustard in lower gangetic plains of West Bengal. <https://www.entomoljournal.com/archives/2020/vol8issue6/PartS/8-6-101-545.pdf>

## **Miscellaneous**

Coombs, A. R. (2020). PhD thesis. Beyond the Nitrogen Thesis: Temporal and Spatial Patterns of *Bras-sica Napus* Oilseed Design Diversification for Crop Rotation, Soil Pest Suppression, Livestock Management, and Convertible Husbandry in the British Agricultural Revolution (s), 1715-1830. <https://knowledge.uchicago.edu/record/2676>

L. Lingmin and H. Jing, "A Vision Method for Rapeseed Amount Measuring," 2020 5th International Conference on Control, Robotics and Cybernetics (CRC), Wuhan, China, 2020, pp. 207-210, <https://doi.org/10.1109/CRC51253.2020.9253481>

Yang, Z.T., Lu, D.X., Hong, EK. et al. Extraction and Separation of Sinapine from Rapeseed Cake and the Mode of Action of Melanin Production Inhibition. *Mol Biol* 54, 911–918 (2020). <https://doi.org/10.1134/S002689332005012X>

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